

**Review Article****Negative Effects of Aflatoxin B1 on Sperm***Aflatoksin B1'in Sperm Üzerine Negatif Etkileri***Veysel DOĞAN***

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Effects of aflatoxin B1 on Sperm
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10.5281/zenodo.6835093**Abstract**

Aflatoxins are secondary metabolites that produced by *Aspergillus* fungi which are soilborne and involved in the decomposition of plant materials in nature. *Aspergillus* species find opportunity to produce aflatoxins in high humidity and temperature conditions such as tropical and subtropical environment. Therefore, in changing climatic conditions caused by global warming, geographical distribution of these fungi to have changed day by day, and find out an opportunity to grow in different feed materials. Aflatoxin B1 (AFB1) is the most toxigenic mycotoxin in these group and threatens both human and animal health due to its carcinogenic and mutagenic properties. In cattle, commonly known adverse effects of AFB1 depending on the chronic exposure such as decreasing milk production and quality, reducing the feed efficiency in beef cattle, increase susceptibility to diseases following suppression of the immune system, compromise ruminal functions, disruption of ruminal microorganism growth and functions.

In this study, the negative effects of AFB1 on bull sperm has been examined. Topics such as negative effects of AFB1 on sperm proteomes, increase in the reactive oxygen species, changes in sperm DNA and plasma membrane integrity are discussed.

Key Words: Aflatoxin, Aflatoxin B1, Sperm**ÖZ**

Aflatoksinler, toprak kaynaklı olan ve bitkisel materyallerin çürümesinde görev alan *Aspergillus* türü mantarlar tarafından üretilen sekonder metabolitlerdir. *Aspergillus* türleri, tropikal ve subtropikal ortamlar gibi yüksek nem ve sıcaklık koşullarında aflatoksin üretme fırsatı bulmaktadır. Bu nedenle, küresel ısınmanın neden olduğu iklim değişikliklerine bağlı olarak aflatoksin üreten mantarların coğrafi yayılışları değişmekte ve farklı yem materyallerinde üreme imkanı bulmaktadırlar. Aflatoksinler içerisinde en toksik olanı aflatoksin B1'dir (AFB1) ve karsinogenik ve mutajenik özelliğinden dolayı insan ve hayvan sağlığını tehdit etmektedir. Aflatoksin B1'e uzun süre maruz kalmaya bağlı olarak sığırlarda; süt veriminde ve kalitesinde azalma, besi sığırlarında yemden yararlanmada düşme, immun sistemin baskılanması ve buna bağlı olarak hastalıklara karşı duyarlılıkta artış, karaciğer fonksiyonlarının baskılanması, rumen fonksiyonlarında azalma, rumen mikroorganizmalarının gelişme ve fonksiyonlarının sekteye uğratılması gibi etkileri yaygın olarak bilinmektedir.

Bu çalışmada, AFB1'in boğa spermi üzerindeki olumsuz etkileri incelenmiştir. Boğa spermindeki proteomlar, reaktif oksijen türlerindeki artışlar ve plazma membran bütünlüğündeki değişiklikler, sperm DNA'sında meydana gelen değişiklikler gibi konular ele alınmıştır.

Anahtar Kelime: Aflatoksin, Aflatoksin B1, Sperm

Highlights

- AFB1 impairs the spermatozoa acrosome reaction and capacitation.
- AFB1 reduces PARK7 expression in spermatozoa and increases ROS generation.
- DNA fragmentation in spermatozoa is increased in males exposed to AFB1.

Introduction

Aflatoxin contamination in various feed materials such as corn, maize, sorghum, rice and wheat are common worldwide (1-3). Aflatoxin contamination of corns cause economic losses in U.S. corn industry between US\$52.1 and US\$1.68 billion (4). Estimates of the economic losses caused by mycotoxin contamination vary, nevertheless, neither of these estimates contain human health impact of aflatoxin contamination.

Aflatoxins are the secondary metabolites produced by fungi, *Aspergillus flavus*, *A. parasiticus* and *Penicillium* species (5-7). Aflatoxins consist four main compounds called Aflatoxin (AF) B1, B2, G1 and G2 (6). Aflatoxin M1 and M2 which are found in milk and dairy products are hydroxylated forms of AFB1 and AFB2, respectively (8). According to the International Agency for Research on Cancer, AFB1 is classified as Group 1 carcinogen to humans (9).

In cattle, long term exposure to aflatoxins can reduce production performance, interrupt liver function, increase the susceptibility to diseases following suppression of immune function (10). The negative effects of aflatoxins such as production performance have been observed by researchers using pure aflatoxins because it allows control of the dose applied and more contamination prevention. However, in nature, different fungi species can grow in the feedstuffs depending on the environmental conditions. Therefore, mycotoxins may cause more severe damage due to the synergistic or additive effects of different types of mycotoxin as well as other metabolites and their fungal sources (11-13). Applebaum et al (1982) observed that the pure AFB1 administration to dairy cows did not affect the milk productions whereas impure AFB1 administration reduced the milk production in dairy cows (14). Due to the synergistic or additive effects of different mycotoxin types, or aflatoxins, may exert their deleterious effects more severely in vivo or in vitro.

Aflatoxin B1 may have toxic to the male reproductive system in animal as well as human. The negative effects of AFB1 on male reproductive system are sorted as pathological changes in testis and epididymis, decreases in the number of leydig cells, and in the number of spermatogenesis, spermatocytes and spermatids (15). In addition, AFB1 exerts its negative effects on human are reported as poor sperm quality and infertility (16).

Effect on sperm capacitation and acrosome reaction

The acrosome reaction is a prerequisite process of spermatozoa for fertilization. Acrosome-reacted spermatozoa are being capable to pass through the zona pellucida subsequently bind the oocyte plasma membrane and fuse with the oocyte (17). All mammalian spermatozoa including human undergo a series changes during their ascent in the female reproductive tract, is called capacitation (18). The acrosome reaction requires to release of hyaluronidase and acrosin enzymes (17). Ataman et al., (2014) have observed significant increase in semen hyaluronidase activity in rams exposed to aflatoxin from 3 week of the trial (19). Researchers had discussed on increase in semen hyaluronidase activity could be arise from an increase in the rate of abnormal or nonviable spermatozoa or could be explained by the transfer of hyaluronidase from the serum into the seminal plasma as a result of chronic intoxication. In contrast, chronic exposure to AFB1 of mammals in dose dependent manner cause low serum testosterone (20, 21). In another hand, the high testosterone levels in sheep results in high serum hyaluronidase levels (22). Therefore, the increase in hyaluronidase activity in semen could not be related to the transfer from the serum into the seminal plasma.

Komsky-Elbaz et al., (2018) reported that AFB1 causes significant decrease the proportion of sperm that reacts to Ca^{++} ionophore and underwent induced acrosome reaction in sperm obtained from the epididymis tail (23). In another study, the ubiquitin-proteosome systems (UPS) that involved in capacitation, acrosome reaction and zona pellucida penetration, have been found affected by the AFB1 (24).

Effect on mitochondria

Alterations in mitochondrial functions caused by the environmental substrates such as aflatoxins are associated with dysfunction male and female infertility (25). The mitochondria involve in ATP synthesis, reactive oxygen species (ROS) production, calcium signaling and apoptosis. Impairment of mitochondrial functions in cells exposed to AFB1 induce apoptosis following activating ROS generation by the mitochondria (29). Komsky et al., (2018) observed that AFB1 induced alterations in mitochondrial membrane potential in spermatozoa (23). In addition, AFB1 reduced expression of PARK7, a protein involved in cell protection against mitochondrial damage and high levels of ROS generation (24, 26). These results suggest that AFB1 reduces fertilization rate via mitochondrial damage. The ubiquitin C-terminal hydrolase L3 protein is located in the mitochondrial sheath and shows reduced expression in male infertility (24).

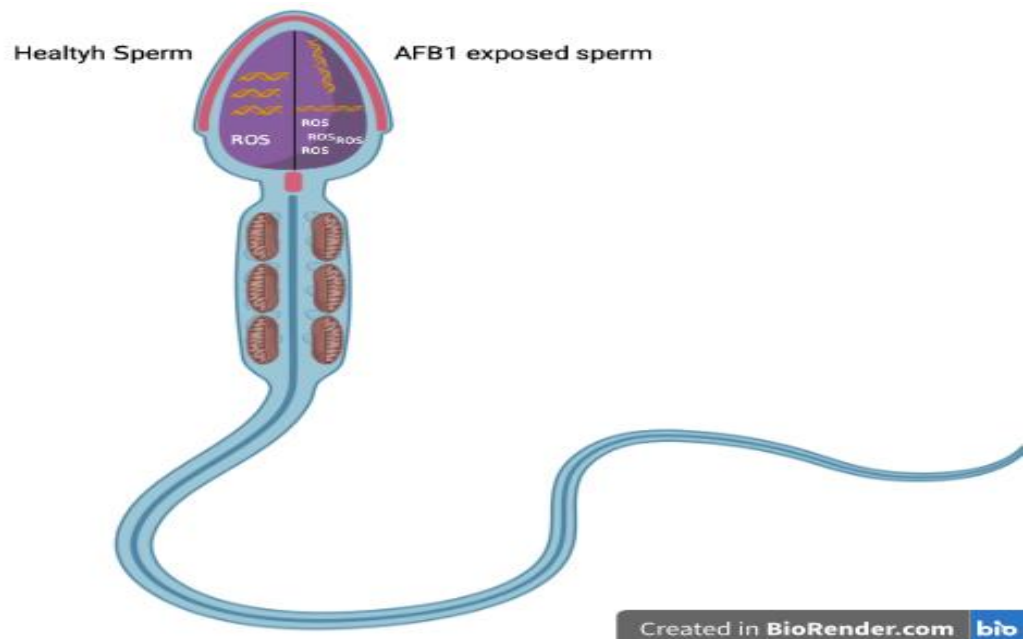


Figure 1: Effects of Aflatoxin B1 (AFB1) on sperm quality. Acrosome reaction is impaired, and apoptosis is induced following activated ROS generation. DNA fragmentation is increased.

DNA Fragmentation

Paternal components which are believed to be crucial for oocyte activation and zygote formation, are delivered by spermatozoon. During fertilization, the proportion of sperm with DNA fragmentation is considered a practical parameter for characterizing semen quality (27, 28). DNA fragmentation reduces sperm fertilization and also effects embryonic development (29), but embryo can repair DNA damage of sperm origin. This process starts after fertilization (30). Komsky et al., (2018) reported DNA fragmentation in bull sperm exposed to AFB1 but no difference on the blastocyst formation between AFB1-treated group and control (23).

Cell membrane integrity is important on the cell viability. Komsky et al., (2018) reported that exposure of low concentrations of AFB1 (1 or 10 μM) reduced the viability of sperm (23). Ram spermatozoa exposed to AFB1 induced the higher rate of dead spermatozoa proportion in comparison with control group (19).

In conclusion, while aflatoxins are considered one of the risk factors for decrease in production performance such as milk production, feed efficiency in dairy and beef cattle, according to the data from the limited studies, they may also accepted as risk factor for male reproduction. These risk factors for male reproduction include decreasing sperm motility and viability, increasing the rate of fragmented DNA, and ROS in spermatozoa. Further studies are required for negative effects on male reproduction exposed to aflatoxins either alone or combined with other mycotoxins.

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References

1. Schenzel J, Forrer HR, Vogelgsang S, et al. Mycotoxin in the environment: I. production and emission from an agricultural test field. *Environ Sci Technol.* 2012; 46:13067-75.
2. Hell K, Cardwell K, Setamou M, et al. The influence of storage practices on aflatoxin contamination in maize in four agroecological zones of Benin, west Africa. *J Stored Prod Res.* 2000; 36:365-82.
3. Kosicki R, Blajet-Kosicka A, Grajweski J, et al. Multiannual mycotoxin survey in feed materials and feedingstuffs. *Anim Feed Sci Technol.* 2016; 215:165-80.
4. Mitchell NJ, Bowers E, Hurburgh C, et al. Potential economic losses to the US corn industry from aflatoxin contamination. *Food Addit Contam Part A.* 2016; 33:540-550.
5. Kew MC. Aflatoxins as a cause of hepatocellular carcinoma. *J Gastrointestin Liver Dis.* 2013; 22:305-10.

6. Zhang D, Li P, Zhang Q, et al. Ultrasensitive nanogold probe-based immunochromatographic assay for simultaneous detection of total aflatoxins in peanuts. *Biosens Bioelectron.* 2011; 26:2877-82.
7. Frisvad JC, Skouboe P, Samson RA. Taxonomic comparison of three different groups of aflatoxin producers and a new effluvin producer of aflatoxin B1, sterigmatocystin and 3-O-methylsterigmatocystin, *Aspergillus rambellii* sp. nov. *Syst Appl Microbiol.* 2005; 28:442-53.
8. Sheng YJ, Eremin S, Mi TJ, et al. The development of a fluorescence polarization immunoassay for aflatoxin detection. *Biomed Environ. Sci.* 2014; 27:126-9.
9. International Agency for Research on Cancer (IARC). Some traditional herbal medicines, some mycotoxins, naphthalene and styrene: Summary of data reported and evaluation. In *IARC Monographs on the Evaluation of the Carcinogenic Risk to Humans*; IARC: Lyon, France. 2002;82: 36.
10. Fink-Gremmels J. The role of mycotoxins in the health and performance of dairy cows. *Veter J.* 2008; 176:84-92.
11. Alassane-Kpembi I, Schatmayr G, Taranu I, et al. 2017. Mycotoxins co-contamination: Methodological aspects and biological relevance of combined toxicity studies. *Crit Rev Food Sci Nutr.* 2017; 57:3489-507.
12. Segvis Klaric M. Adverse effects of combined mycotoxins. *Arh Hig Rada Toksikol.* 2012;63: 519-30.
13. Segvic Klaric M, Rasic D, Peraica M. Deleterious effects of mycotoxin combinations involving ochratoxin A. *Toxins.* 2013; 5:1965-87.
14. Applebaum RS, Brackett RW, Wiseman DW, et al. Responses of dairy cows to dietary aflatoxin: feed intake and yield, toxin content, and quality of milk of cows treated with pure and impure aflatoxin. *J Dairy Sci.* 1982; 65:1503-08.
15. Murad AF, Ahmed S, Aboad S. Toxicity effect of Aflatoxin B1 on reproductive system of albino male rats. *Pak J Bio Sci.* 2015; 18:107-14.
16. Eze UA, Routledge MN, Okonofua FE, et al. Mycotoxin exposure and adverse reproductive health outcomes in Africa: a review. *World Mycotoxin Journal.* 2018; 11:321-39.
17. Yanagimachi R. Mammalian fertilization. In: Knobil E, Neill JD (Eds.) *The physiology of reproduction.* Vol. 2. Raven Press, New York, 1994.
18. Cornwall GA. 2014. Role of posttranslational protein modifications in epididymal sperm maturation and extracellular quality control. *Adv Exp Med Biol.* 2014; 759:159-80.
19. Ataman MB, Donmez HH, Donmez N, et al. Protective effect of esterified glucomannan on aflatoxin-induced changes in testicular function, sperm quality, and seminal plasma biochemistry in rams. *Theriogenology*, 2014;81:373-80.
20. Supriya C, Girish BP, Reddy PS. Aflatoxin B1-induced reproductive toxicity in male rats: Possible mechanism of action. *Int J Toxicol.* 2014; 33:155-61.
21. Salem MH, Kamel KI, Yousef MI, et al. Protective role of ascorbic acid to enhance semen quality of rabbits treated with sublethal doses of aflatoxin B1. *Toxicology*, 2001;162:209-18.
22. Tanyıldızı S. 2002. Effects of progesterone and testosterone on the hyaluronidase activities and sperm characteristics in sheep. *Turk J Vet Anim Sci.* 2002; 26:1137-43.
23. Komsky-Elbaz A, Saksier M, Roth Z. Aflatoxin B1 impairs sperm quality and fertilization competence. *Toxicology.* 2018; 393:42-50.
24. Komsky-Elbaz A, Kalo D, Roth Z. Effect of aflatoxin B1 on bovine spermatozoa's proteome and embryo's transcriptome. *Reproduction.* 2020; 160:709-23.
25. Ramalho-Santos J, Varum S, Amaral S, et al. Mitochondrial functionality in reproduction: from gonads and gametes to embryos and embryonic stem cells. *Hum Reprod Update.* 2009; 15:553-72.
26. Sun Y, Zhang WJ, Zhao X, et al. PARK7 protein translocating into spermatozoa mitochondria in Chinese asthenozoospermia. *Reproduction*, 2014;148:249-57.
27. Sergerie M, Lafors G, Bujan L, et al. Sperm DNA fragmentation: threshold value in male fertility. *Hum Reprod.* 2005; 20:3446-51.
28. Dogan S, Vargovic P, Oliveria R, et al. Sperm protamine-status correlates to the fertility of breeding bulls. *Biol Reprod.* 2015; 92:92.
29. Ioannou D, Miller D, Griffin DK, et al. Impact of sperm DNA chromatin in the clinic. *J Assist Reprod Genet.* 2016; 33:157-66.
30. Uppangala S, Pudakalakatti S, D'souza F, et al. Influence of sperm DNA damage on human preimplantation embryo metabolism. *Reprod Biol.* 2016; 16:234-41.